



Integration of Chip-based Sample Preconcentration into the μ ChemLab™/CB Analysis Platform

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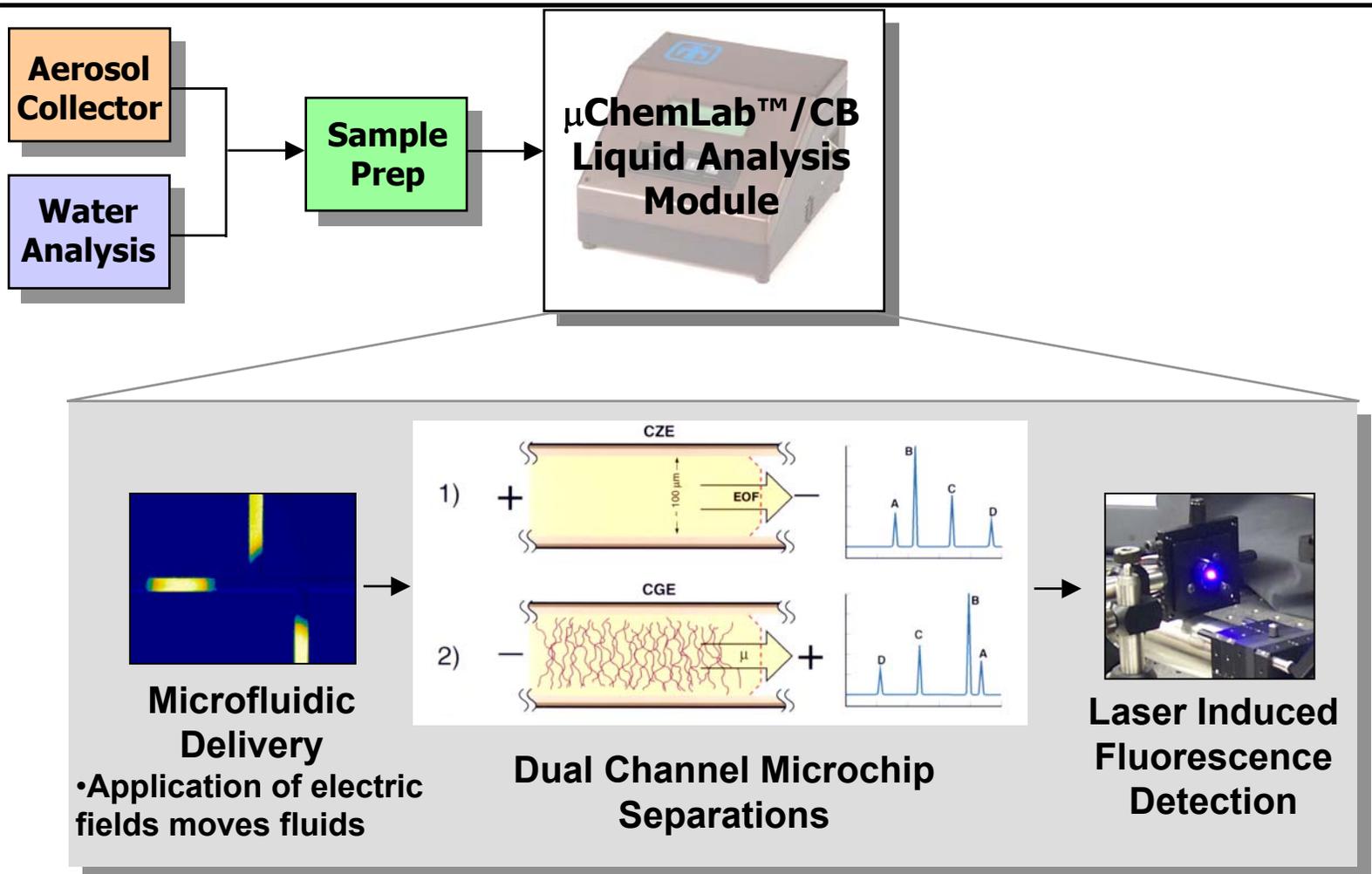
HPCE 2003
San Diego, CA



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μ ChemLab™ Separation Platform





μ ChemLab™ Separation Platform

Two Channel Device

Capillary Gel Electrophoresis (CGE) Channel

Beckman 14-200 SDS Gel

Reversed polarity

Capillary Zone Electrophoresis (CZE) Channel

10 mM phytic acid, pH 9.5 containing 2 mM DAPS (zwitterionic detergent)

Normal polarity

Fluorescamine labeling

Fluorogenic dye

Fast-reacting, amine specific

Ex/Em 390 nm/480nm

Detection limits

nM for CGE



Approaches to Preconcentration

- **Solid Phase Extraction** (on-going efforts for the miniaturization of SPE)
- **Electrokinetic Trapping (P411-T: Novel Miniaturized Protein Preconcentrator Based On Electrokinetic Trapping, Anup K. Singh; Daniel J. Throckmorton; Brian J. Kirby)**

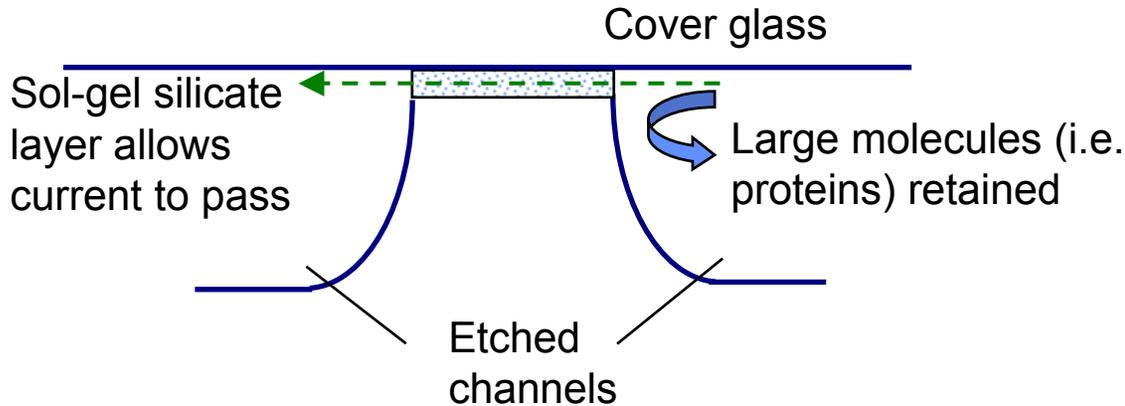
- **Salt-Bridge**

[1] J. Khandurina, T.E. McKnight, S.C. Jacobson, L.C. Waters, R.S. Foote, J.M. Ramsey, *Analytical Chemistry* 72 (2000) 2995-3000

[2] R. S. Foote, J. Khandurina, S. C. Jacobson, J. M. Ramsey, Preconcentration of Proteins on Microchips for Enhanced Detection, Poster presentation at HPCE 2001

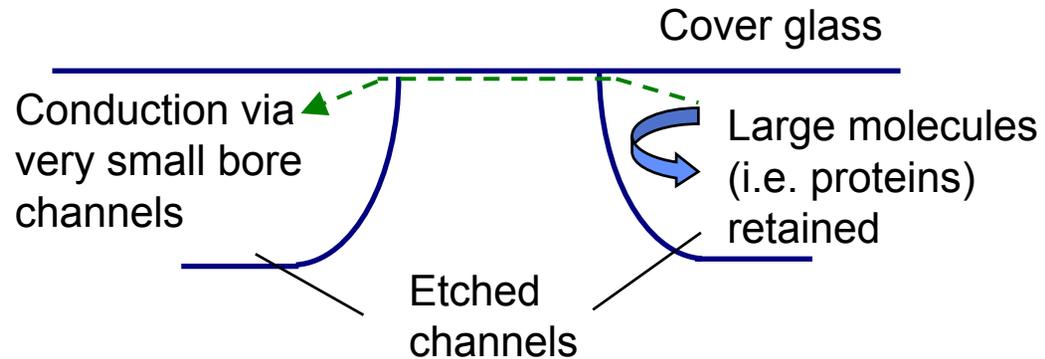
Integration into the μ ChemLabTM/CB

Silicate layer Approach:



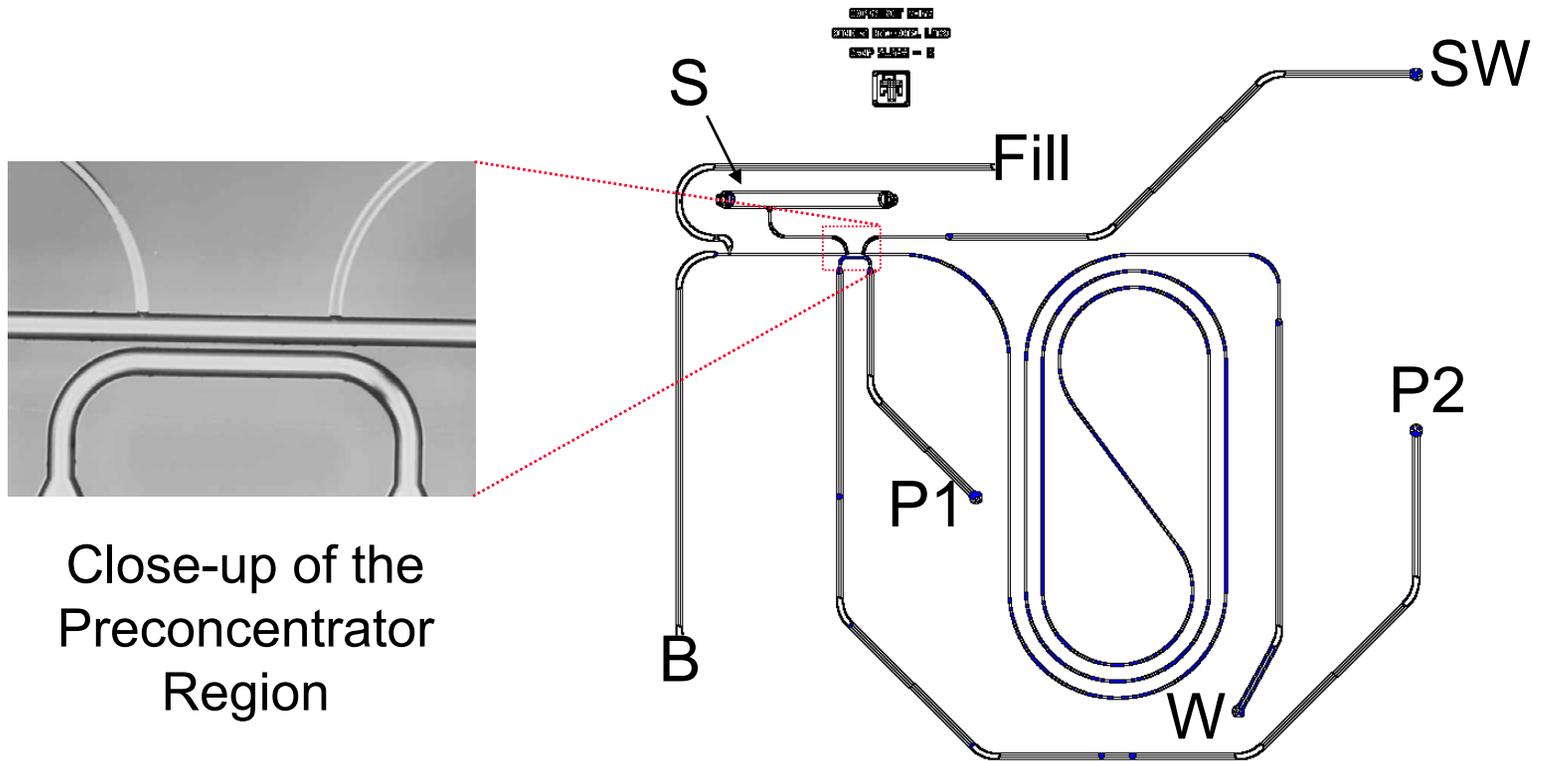
Alternative Approach:

- Eliminate silicate layer
- Makes use of narrow gap and natural surface roughness



▪ Jean Pierre Alaire, Stephen C. Jacobson, B. Scott Broyles, Timothy E. McKnight, Christopher T. Culbertson and J. Michael Ramsey, Electroosmotically Induced Hydraulic Pumping on Microchips. *μ TAS 2001*. J. Michael Ramsey and A. van den Berg. ed.s, (Kluwer Academic, Amsterdam), (2001), pp. 127-128.

Chip Layout





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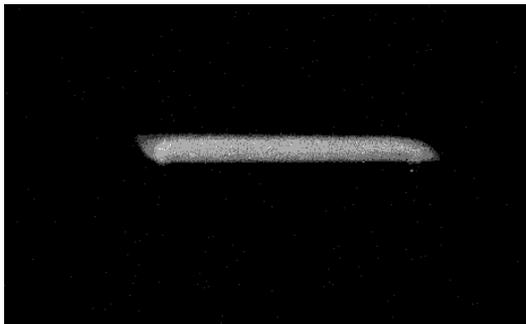
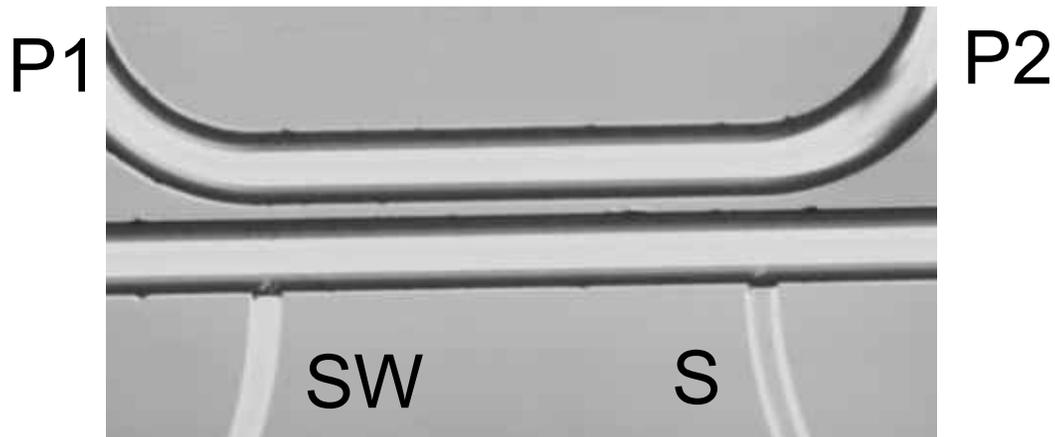
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Detection limits

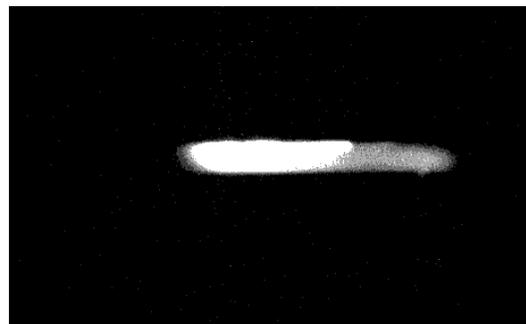
nM for CGE

Imaging Normal and Preconcentrated Injections in CGE



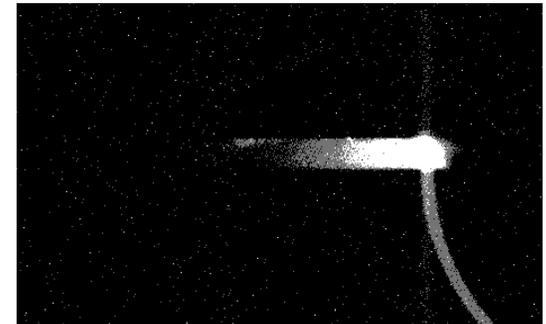
Normal Mode

S → SW



P1 Preconcentration
Mode

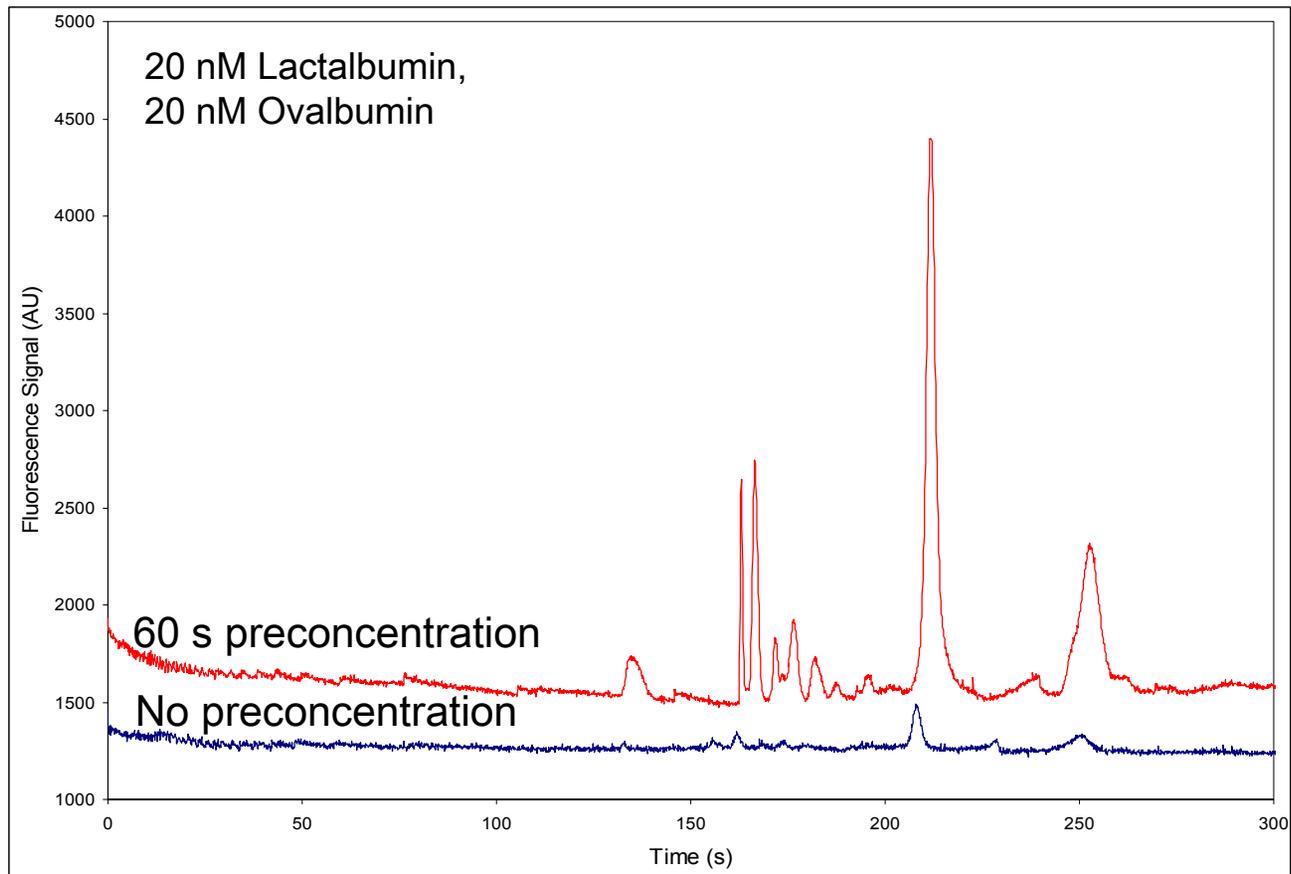
S → P1



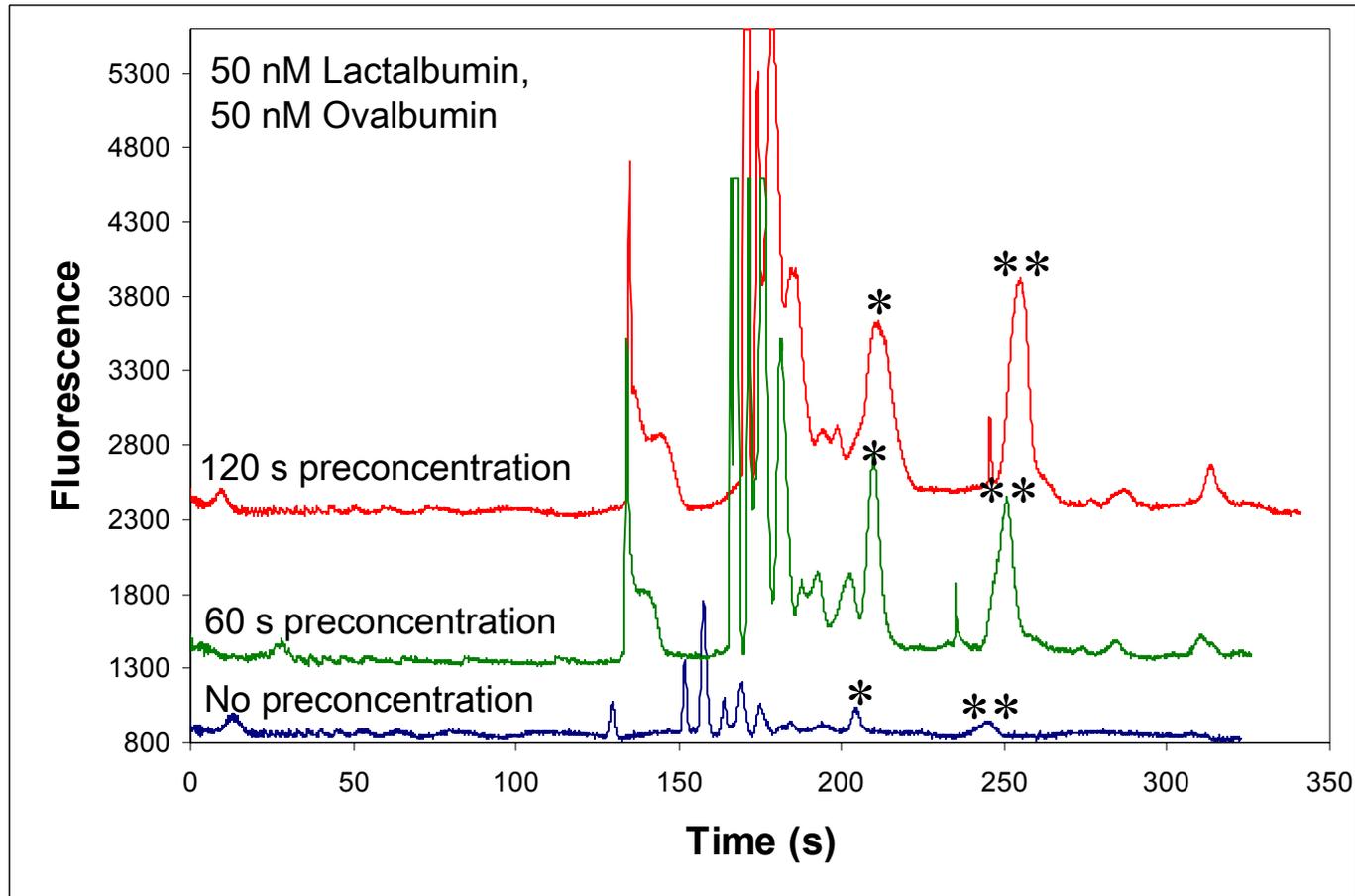
P2 Preconcentration
Mode

S → P2

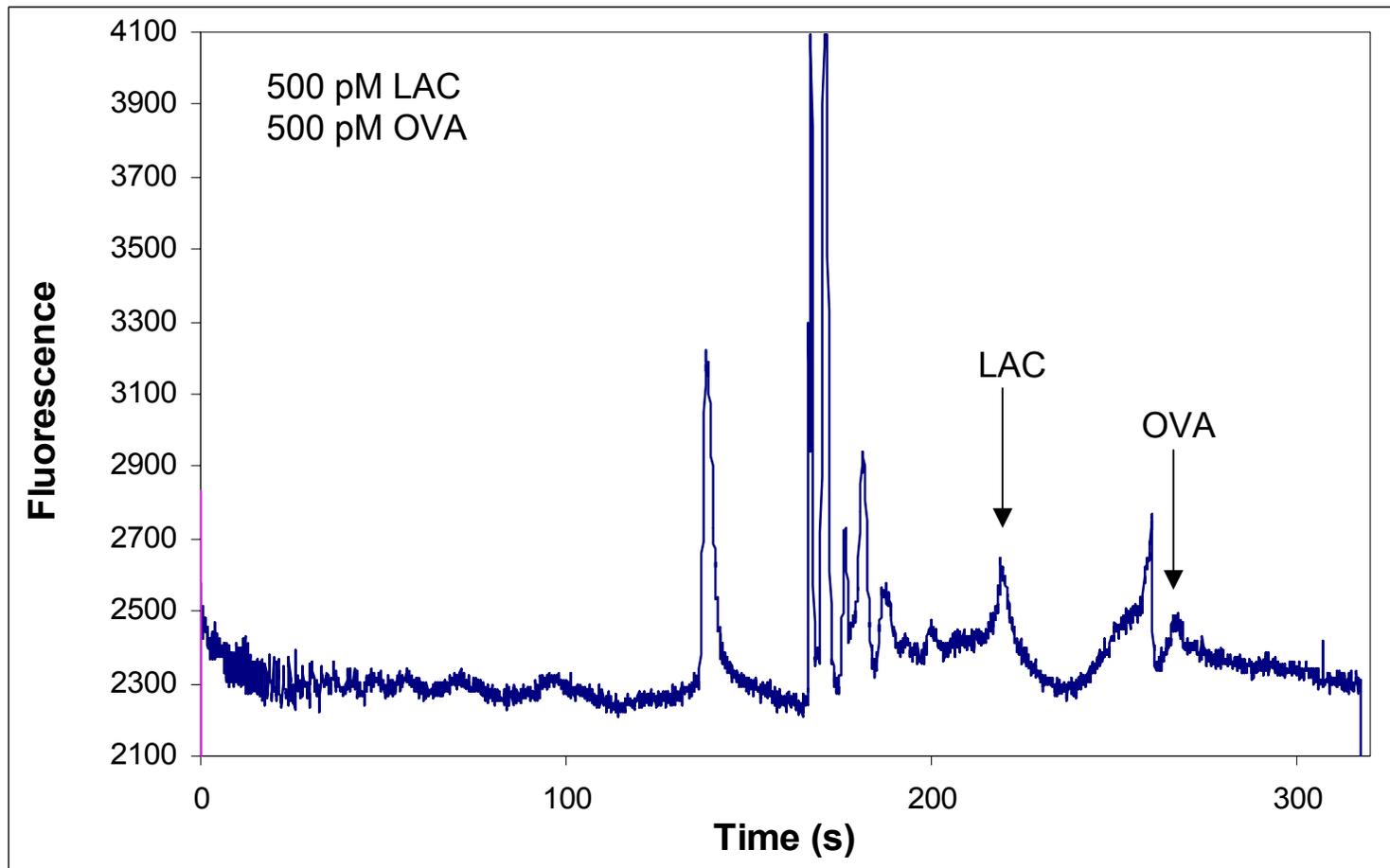
CGE Separations Using Normal and Preconcentrated Injections



Peak Areas in CGE Separations Increase as a Function of Preconcentration Duration



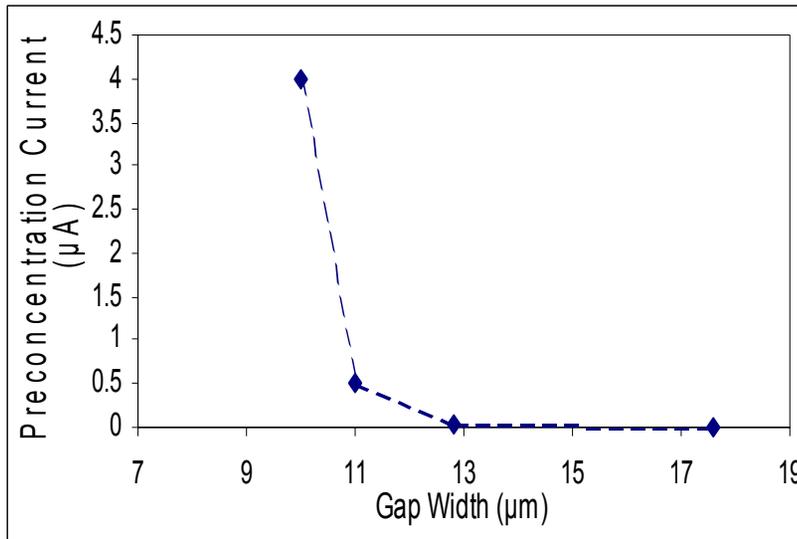
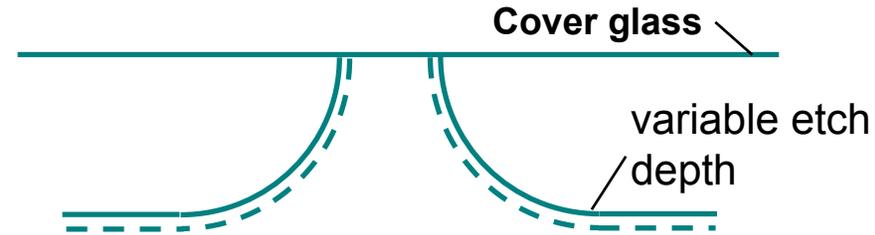
Sub-Nanomolar Detection Limits Using CGE Separations and Preconcentration



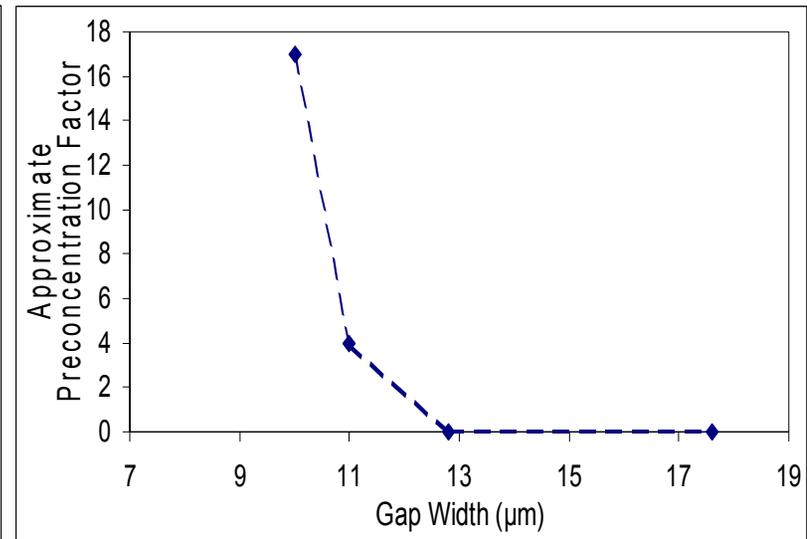
Characterization of Performance as a Function of Gap Width

Gap width

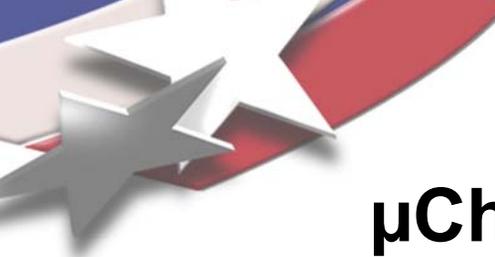
- Isotropic etch depth used to generate gaps of different widths
- For comparison purposes, all preconcentrations were held to 60s



*Single Batch Data



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Preconcentration Using CZE

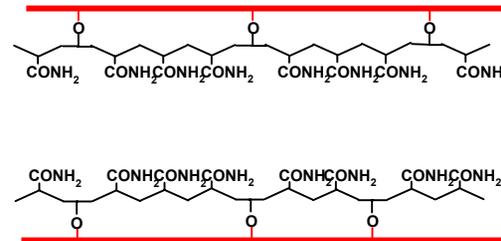
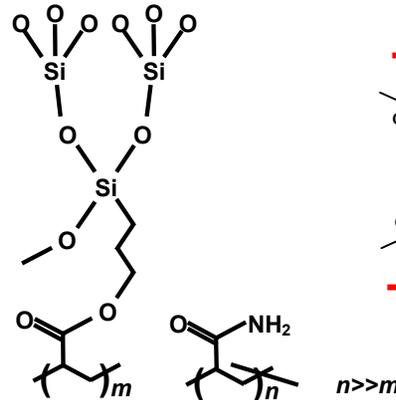
Elimination of EOF is required

- no bulk flow of fluid through the gap
∴ residual EOF could lead to pressure generation

Coating with linear polyacrylamide

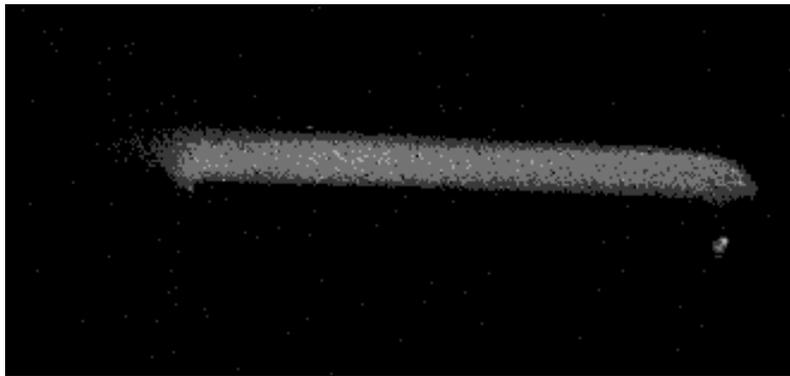
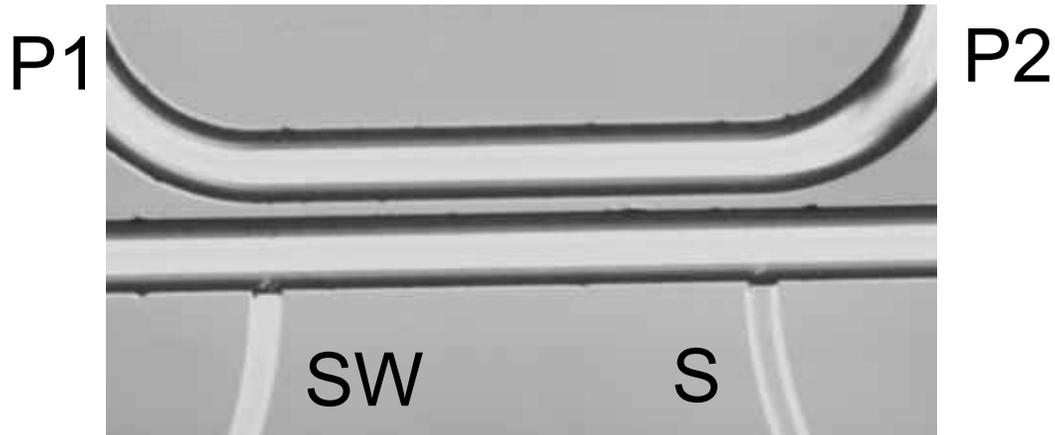
SAM application

UV light polymerization

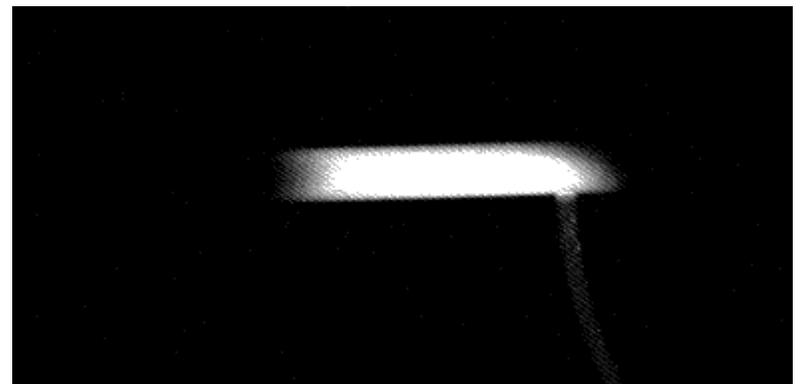


•Julie Fruetel, Victoria VanderNoot, Jay West, Brian Kirby, Ernest Hasselbrink and Timothy Shepodd, Laser-polymerized thin-film coating for protein analysis by CGE in a microchip, HPCE 2002

Imaging Normal and Preconcentrated Injections in CZE

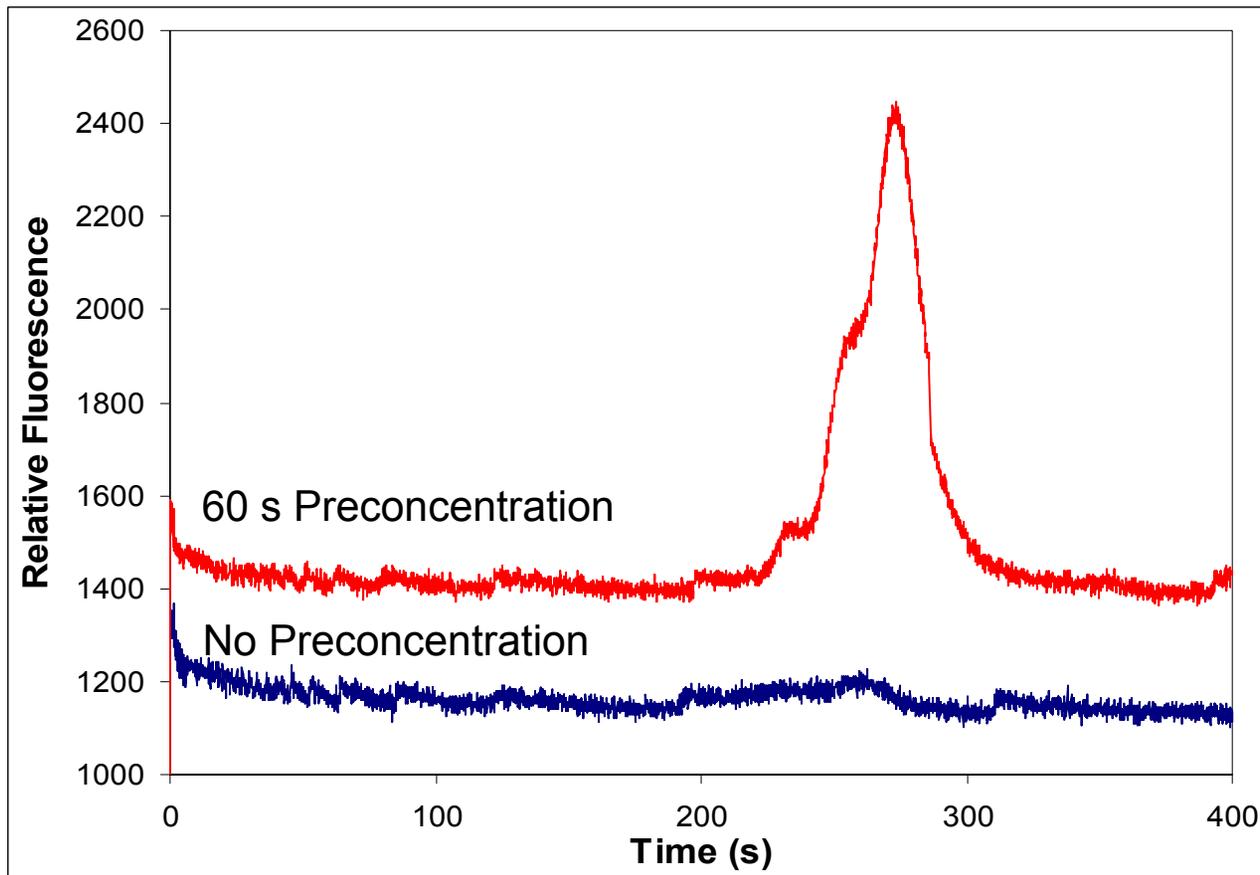


Normal Mode
 $S \rightarrow SW$



P1 Preconcentration Mode
 $S \rightarrow P1$

CZE Separations Using Normal and Preconcentrated Injections



Running buffer: 5 mM phosphate, pH 8 containing 5 mM CAS-U zwitterionic detergent
Reverse polarity



Summary

- Preconcentration has been incorporated in the chip design with no additional processing steps during chip fabrication
- Successfully demonstrated with both CGE and CZE
 - Preconcentration factors of 10-20x are routinely achievable in only 60s
 - Coating and/or elimination of EOF is essential for CZE separations
 - Buffer conditions in CZE will need to be optimized to achieve both good separation efficiency and the elimination of EOF



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The μ ChemLab Technical Team

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Yolanda Fintschenko	George Sartor	Boyd Wiedenman
Julie Fruetel	George Schubert	Dan Yee

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